

1. General Information

Cohort ID	5003_20
Title (Study Name)	COVID-19 Convalescent Cohort
Principal investigator	Prof. Robert Krause, Dr. Christian Gölly, Prof. Peter Schlenke, Prof. Ivo Steinmetz
Contact information	pm-biobank@medunigraz.at
Funding agency	

2. Description

This longitudinal cohort comprise a collection of samples from 364 convalescents who were infected with SARS-CoV-2.

- Sub cohort I: persons who were officially tested positive for SARS-CoV-2 by RT-PCR between 29th of February 2020 and 15th of January 2021, showed symptoms of COVID-19 and have recovered. (342 participants)

- Sub cohort II: persons recovered from suspected COVID-19 infection (but COVID-19 infection has not been confirmed by RT-PCR) (22 participants)

Study design: Participants were invited 5 times: 1.visit and 4 follow-up visits were performed 1, 2, 5 and 12 months after the first visit where the samples were collected. During the first visit, participants filled in a questionnaire concerning symptoms, comorbidities, prehistory and lifestyle.

This cohort serves for e.g.:

- the development and validation of new antibody tests and neutralization assays
- the investigation of antibody titres over time
- a better characterization of the course of COVID-19
- the identification of diagnostic or prognostic biomarkers

3. Details

ICD 10/O codes / Healthy	Healthy	
Key words	COVID-19 recovered patients, SARS-Cov-2, PCR test, antibody test, longitudinal cohort	
Collection / Cohort size 12/2022	38.115 aliquots from 364 participants (5 visits)	
Informed Consent (IC)	<input checked="" type="checkbox"/> Broad Biobank IC <input checked="" type="checkbox"/> Specific Study IC	
Status	<input type="checkbox"/> In progress / compl. date: 03/2022 <input checked="" type="checkbox"/> Completed	
Inclusion criteria	Age distribution	18-81 years
	Sex distribution (f:m)	61:39
	Others	Patients who have recovered from COVID-19
Earliest access	As of now	

Quality-standards	<input checked="" type="checkbox"/> ISO 9001:2015 (SOPs)
Associated publications / references	<p><u>Long-lasting immune response to a mild course of PCR-confirmed SARS-CoV-2 infection: A cohort study</u> Kral S, Banfi C, Niedrist T, Sareban N, Guelly C, Kriegl L, Schiffmann S, Zurl C, Herrmann M, Steinmetz I, Schlenke P, Berghold A, Krause R. https://pubmed.ncbi.nlm.nih.gov/34433071/</p> <p><u>Longitudinal comparison of automated SARS-CoV-2 serology assays in assessing virus neutralization capacity in COVID-19 convalescent sera</u> Niedrist T, Drexler C, Torreiter PP, Matejka J, Strahlhofer-Augsten M, Kral S, Riegler S, Güll C, Zurl C, Kriegl L, Krause R, Berghold A, Steinmetz I, Schlenke P, Herrmann M. https://pubmed.ncbi.nlm.nih.gov/35085385/</p> <p><u>Lower Levels of ABO Anti-A and Anti-B of IgM, IgG and IgA Isotypes in the Serum but Not the Saliva of COVID-19 Convalescents.</u> Matzhold EM, Körmöcz GF, Banfi C, Schönbacher M, Drexler-Helmberg C, Steinmetz I, Berghold A, Schlenke P, Wagner GE, Stoissner A, Kleinhappl B, Mayr WR, Wagner T https://pubmed.ncbi.nlm.nih.gov/35956128/</p> <p><u>Preanalytical stability of SARS-CoV-2 anti-nucleocapsid antibodies</u> Niedrist T, Kriegl L, Zurl CJ, Schmidt F, Perkmann-Nagele N, Mucher P, Repl M, Flieder I, Radakovics A, Sieghart D, Radner H, Aletaha D, Binder CJ, Güll C, Krause R, Herrmann M, Wagner OF, Perkmann T, Haslacher H. https://pubmed.ncbi.nlm.nih.gov/36323338/</p> <p><u>Development of a Rapid Live SARS-CoV-2 Neutralization Assay Based on a qPCR Readout.</u> Lichtenegger S, Saiger S, Hardt M, Kulnik S, Wagner GE, Kleinhappl B, Assig K, Zauner A, Ober M, Kimpel J, von Laer D, Zatloukal K, Steinmetz I. https://pubmed.ncbi.nlm.nih.gov/35642515/</p> <p><u>SARS-CoV-2 neutralizing activity of polyphenols in a special green tea extract preparation.</u> Kicker E, Tittel G, Schaller T, Pferschy-Wenzig EM, Zatloukal K, Bauer R. https://pubmed.ncbi.nlm.nih.gov/35144138/</p>

4. Material available (aliquot size) and storage conditions

Material	<input checked="" type="checkbox"/> Serum (235 µl)	<input checked="" type="checkbox"/> -80°C
	<input checked="" type="checkbox"/> EDTA plasma (300 µl)	<input checked="" type="checkbox"/> -80°C
	<input checked="" type="checkbox"/> EDTA Buffy coat (580 µl)	<input checked="" type="checkbox"/> -80°C
	<input checked="" type="checkbox"/> Li-Hep plasma (235 µl)	<input checked="" type="checkbox"/> -80°C
	<input checked="" type="checkbox"/> Na-Citrate plasma (235 µl)	<input checked="" type="checkbox"/> -80°C
	<input checked="" type="checkbox"/> Saliva	<input checked="" type="checkbox"/> -80°C
	<input checked="" type="checkbox"/> Nasopharyngeal swab	<input checked="" type="checkbox"/> -80°C
	<input checked="" type="checkbox"/> DNA	<input checked="" type="checkbox"/> -80°C

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