


	<b>O-FIS Qualitätsmanagementsystem</b> Formblatt <b>Collection and Cohort Profile</b>	CL312 Seite 1 von 3
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## 1. General Information

<b>Cohort ID</b>	5003_20
<b>Title (Study Name)</b>	COVID-19 Convalescent Cohort
<b>Principal investigator</b>	Prof. Robert Krause, Dr. Christian Güllly, Prof. Peter Schlenke, Prof. Ivo Steinmetz
<b>Contact information</b>	<a href="mailto:pm-biobank@medunigraz.at">pm-biobank@medunigraz.at</a>
<b>Funding agency</b>	

## 2. Description

<p>This longitudinal cohort comprise a collection of samples from 364 convalescents who were infected with SARS-CoV-2.</p> <ul style="list-style-type: none"> <li>• Sub cohort I: persons who were officially tested positive for SARS-CoV-2 by RT-PCR between 29th of February 2020 and 15th of January 2021, showed symptoms of COVID-19 and have recovered. (342 participants)</li> <li>• Sub cohort II: persons recovered from suspected COVID-19 infection (but COVID-19 infection has not been confirmed by RT-PCR) (22 participants)</li> </ul> <p>Study design: Participants were invited 5 times: 1.visit and 4 follow-up visits were performed 1, 2, 5 and 12 months after the first visit where the samples were collected. During the first visit, participants filled in a questionnaire concerning symptoms, comorbidities, prehistory and lifestyle.</p> <p>This cohort serves for e.g.:</p> <ul style="list-style-type: none"> <li>• the development and validation of new antibody tests and neutralization assays</li> <li>• the investigation of antibody titres over time</li> <li>• a better characterization of the course of COVID-19</li> <li>• the identification of diagnostic or prognostic biomarkers</li> </ul>
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## 3. Details

<b>ICD 10/O codes / Healthy</b>	Healthy	
<b>Key words</b>	COVID-19 recovered patients, SARS-Cov-2, PCR test, antibody test, longitudinal cohort	
<b>Collection / Cohort size</b> 12/2023	36.665 aliquots from 364 participants (5 visits)	
<b>Informed Consent (IC)</b>	<input checked="" type="checkbox"/> Broad Biobank IC	
	<input checked="" type="checkbox"/> Specific Study IC	
<b>Status</b>	<input type="checkbox"/> In progress / compl. date: 03/2022	
	<input checked="" type="checkbox"/> Completed	
<b>Inclusion criteria</b>	<b>Age distribution</b>	18-81 years
	<b>Sex distribution (f:m)</b>	61:39
	<b>Others</b>	Patients who have recovered from COVID-19
<b>Earliest access</b>	As of now	

<b>Quality-standards</b>	<input checked="" type="checkbox"/> ISO 9001:2015 (SOPs)
<b>Associated publications / references</b>	<p><u>Long-lasting immune response to a mild course of PCR-confirmed SARS-CoV-2 infection: A cohort study</u> Kral S, Banfi C, Niedrist T, Sareban N, Guelly C, Kriegl L, Schiffmann S, Zurl C, Herrmann M, Steinmetz I, Schlenke P, Berghold A, Krause R. <a href="https://pubmed.ncbi.nlm.nih.gov/34433071/">https://pubmed.ncbi.nlm.nih.gov/34433071/</a></p> <p><u>Longitudinal comparison of automated SARS-CoV-2 serology assays in assessing virus neutralization capacity in COVID-19 convalescent sera</u> Niedrist T, Drexler C, Torreiter PP, Matejka J, Strahlhofer-Augsten M, Kral S, Riegler S, Güllly C, Zurl C, Kriegl L, Krause R, Berghold A, Steinmetz I, Schlenke P, Herrmann M. <a href="https://pubmed.ncbi.nlm.nih.gov/35085385/">https://pubmed.ncbi.nlm.nih.gov/35085385/</a></p> <p><u>Lower Levels of ABO Anti-A and Anti-B of IgM, IgG and IgA Isotypes in the Serum but Not the Saliva of COVID-19 Convalescents.</u> Matzhold EM, Körmöczí GF, Banfi C, Schönbacher M, Drexler-Helmsberg C, Steinmetz I, Berghold A, Schlenke P, Wagner GE, Stoisser A, Kleinhapfl B, Mayr WR, Wagner T <a href="https://pubmed.ncbi.nlm.nih.gov/35956128/">https://pubmed.ncbi.nlm.nih.gov/35956128/</a></p> <p><u>Preanalytical stability of SARS-CoV-2 anti-nucleocapsid antibodies</u> Niedrist T, Kriegl L, Zurl CJ, Schmidt F, Perkmann-Nagele N, Mucher P, Repl M, Flieder I, Radakovics A, Sieghart D, Radner H, Aletaha D, Binder CJ, Güllly C, Krause R, Herrmann M, Wagner OF, Perkmann T, Haslacher H. <a href="https://pubmed.ncbi.nlm.nih.gov/36323338/">https://pubmed.ncbi.nlm.nih.gov/36323338/</a></p> <p><u>Development of a Rapid Live SARS-CoV-2 Neutralization Assay Based on a qPCR Readout.</u> Lichtenegger S, Saiger S, Hardt M, Kulnik S, Wagner GE, Kleinhapfl B, Assig K, Zauner A, Ober M, Kimpel J, von Laer D, Zatloukal K, Steinmetz I. <a href="https://pubmed.ncbi.nlm.nih.gov/35642515/">https://pubmed.ncbi.nlm.nih.gov/35642515/</a></p> <p><u>SARS-CoV-2 neutralizing activity of polyphenols in a special green tea extract preparation.</u> Kicker E, Tittel G, Schaller T, Pferschy-Wenzig EM, Zatloukal K, Bauer R. <a href="https://pubmed.ncbi.nlm.nih.gov/35144138/">https://pubmed.ncbi.nlm.nih.gov/35144138/</a></p>

#### 4. Material available (aliquot size) and storage conditions

<b>Material</b>	<input checked="" type="checkbox"/> Serum (235 µl)	<input checked="" type="checkbox"/> -80°C
	<input checked="" type="checkbox"/> EDTA plasma (300 µl)	<input checked="" type="checkbox"/> -80°C
	<input checked="" type="checkbox"/> EDTA Buffy coat (580 µl)	<input checked="" type="checkbox"/> -80°C
	<input checked="" type="checkbox"/> Li-Hep plasma (235 µl)	<input checked="" type="checkbox"/> -80°C
	<input checked="" type="checkbox"/> Na-Citrate plasma (235 µl)	<input checked="" type="checkbox"/> -80°C
	<input checked="" type="checkbox"/> Saliva	<input checked="" type="checkbox"/> -80°C
	<input checked="" type="checkbox"/> Nasopharyngeal swab	<input checked="" type="checkbox"/> -80°C
	<input checked="" type="checkbox"/> DNA	<input checked="" type="checkbox"/> -80°C

**Dokument erstellt** (tt/mm/yyyy):  
11/03/2021

**Letzte inhaltliche Aktualisierung** (tt/mm/yyyy):  
21/03/2024